

Plant-based Diets and Low-Insulin Lifestyles May Suppress Oxidative Stress by Disinhibiting FOXO3a

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ABSTRACT

FOXO transcription factors, notably FOXO3a, promote transcription of a wide range of mitochondrial antioxidant enzymes as well as catalase – an effect thought to contribute to the beneficial impact of caloric restriction on longevity in rodents. Activated Akt suppresses FOXO3a activity by promoting its exclusion from the nucleus. Hence, in the many tissues responsive to IGF-I and/or insulin – hormones which stimulate Akt activity – high circulating levels of free IGF-I or of insulin may have a pro-oxidant impact. Moreover, high insulin levels boost IGF-I bioactivity by inhibiting hepatic production of IGF-BP-1, an IGF-I antagonist. Hence, the combination of a moderate-protein plant-based diet – which down-regulates hepatic IGF-I production via essential amino acid restriction – and a lifestyle that minimizes diurnal insulin levels, would appear likely to offer worthwhile antioxidant protection, possibly contributing to the markedly reduced risk for “Western” cancers and cardiovascular disease characteristic of quasi-vegan societies. The moderate methionine restriction associated with plant-based diets might also aid this effect by diminishing superoxide production via complex I of the mitochondrial respiratory chain, as observed in methionine-restricted rodents.

Growth Factor Suppression of FOXO3a Activity Promotes Oxidative Stress

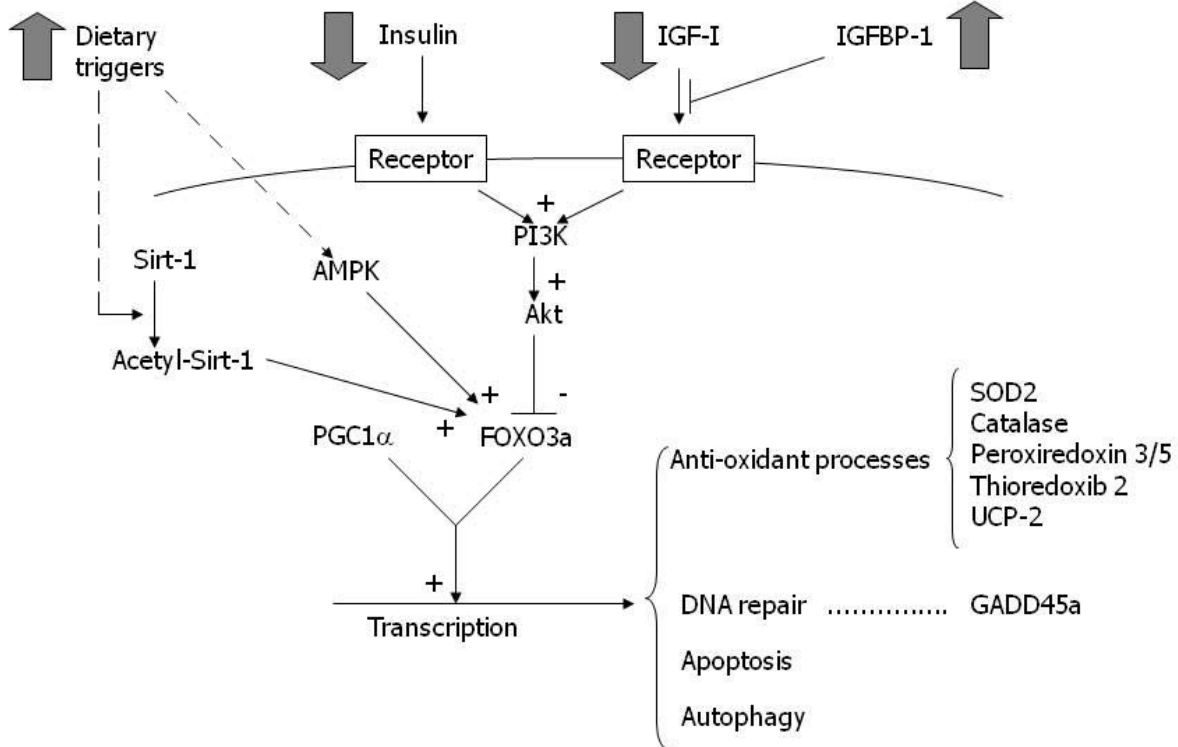
The FOXO transcription factors are homologs of transcription factors found in lower organisms such as worms and flies whose activity is inhibited by signaling pathways homologous to the insulin/IGF-I pathway in vertebrates. Calorie restriction activates these factors in worms and flies, and fails to increase lifespan if these factors are genetically ablated.[1-3] Hence, the FOXO transcription factors are suspected to contribute to lifespan increase in calorically restricted vertebrates. Seemingly consistent with this view are studies demonstrating that certain alleles of the polymorphic FOXO3a gene are significantly enriched in aged humans, presumably because they confer a survival advantage.[4-7]

Control of oxidative stress is clearly necessary, though not likely sufficient, for lifespan extension. Recent research demonstrates that, in human vascular endothelial cells, FOXO3a, acting in conjunction with the transcriptional coactivator PGC-1alpha, boosts

transcription and expression of genes coding for a range of antioxidant enzymes, including the mitochondrial (manganese-dependent) superoxide dismutase, catalase, peroxiredoxin 3, peroxiredoxin 5, thioredoxin 2, thioredoxin reductase 2, as well as for UCP-2 and PGC-1alpha.[8-13] Working in collaboration, these proteins can be expected to suppress mitochondrial release of hydrogen peroxide and peroxynitrite, and to lessen the functional impact of oxidative stress on mitochondrial structure and function; moreover, catalase could also aid control of oxidative stress of cytoplasmic origin. Other key targets of FOXO transcriptional activity include genes that promote efficient DNA repair (GADD45a), apoptosis, autophagy, cell cycle arrest, immune regulation, and muscle atrophy.[14, 15]

Growth factor signaling inhibits the transcriptional activity of the FOXO transcription factors.[14, 16] (See Figure 1.) This comes about because the kinase Akt – activated via PI3K – phosphorylates these factors, thereby promoting their exclusion from the nucleus. (This effect is seen with FOXO1, FOXO3a, and FOXO4, whereas Akt-mediated phosphorylation of FOXO6, expressed primarily in the brain, inhibits

Figure 1



its transcriptional activity without regulating its subcellular location.¹⁷⁾ In the many tissues that are responsive to IGF-I and/or insulin, elevated blood levels of insulin and/or free IGF-I can therefore be expected to impede antioxidant defenses by inhibiting the transcriptional activity of FOXO3a and other FOXO factors. This effect presumably would be somewhat less functionally meaningful in tissues in which autocrine growth factor activity makes an important contribution to Akt activation.

Antioxidant Potential of Plant-based Diets and Low-Insulin Lifestyles

Vegan diets of moderate protein content are associated with reduced circulating levels of IGF-I, likely because a modest degree of essential amino acid restriction suppresses hepatic IGF-I synthesis and secretion.[18-26] Moreover, certain lifestyle strategies are characterized by relatively low diurnal insulin levels, and thus would be likely to lessen Akt

activation both in insulin-sensitive tissues, and – owing to an expected increase in hepatic production of the IGF-I antagonist IGFBP-1[27, 28] – in IGF-I responsive tissues as well. Feasible measures for lessening diurnal insulin secretion include exercise training, leanness, diets with low saturate-unsaturate ratios, reliance on lower-glycemic-index carbohydrates, avoidance of insulinotropic milk protein, calorie restriction, meal-skipping, and carbohydrate-concentrated diets.[29-36] Addition of almonds, soluble fiber, and vinegar to the diet may also diminish diurnal insulin secretion.[37-39] Vegan and Mediterranean diets appear to be inherently useful in this regard, owing to their low saturate-unsaturate ratios and tendencies to promote relative leanness.[29]

It is therefore reasonable to propose that vegan diets of moderate protein content, as well as low-insulin lifestyles, can boost antioxidant protection in many tissues by increasing FOXO activity and consequently the expression of a number of key antioxidant enzymes and proteins. While it has generally been assumed that plant-based diets rich in

antioxidant phytochemicals would promote antioxidant defense, both by the direct scavenging activity of these phytochemicals and their propensity to trigger phase 2 induction,[40-45] the mechanism proposed here is independent of, and presumably complementary to, the protective impact of phytochemical induction.

Vegan diets which are relatively restricted in methionine may also have the potential to decrease mitochondrial oxidant production by an additional mechanism. Significant dietary methionine restriction can increase maximal lifespan in rodents, even with ad libitum calorie consumption; the magnitude of this effect is about half that of optimal calorie restriction.[46-48] Studies by Barja and colleagues have shown that methionine restriction somehow suppresses superoxide production by complex I of the mitochondrial respiratory chain.[49-51] Theoretically, since vegan diets of modest protein content tend to be rather low in methionine, this phenomenon might contribute to antioxidant protection in vegans.[49, 52] It should be noted however, that the diets used to implement methionine restriction in rodent studies have been synthetic diets devoid of cysteine; hence, these diets are sulfur deficient, not just methionine deficient. Whether concurrent ingestion of cysteine might blunt the impact of methionine-restricted diets on longevity and mitochondrial oxidant production needs to be assessed. (To the contrary, it might be argued that cysteine ingestion would amplify the antioxidant merits of such diets by boosting glutathione production.)

In individuals with metabolic syndrome, adoption of diets low in saturated fat (such as vegan or Mediterranean diets typically are) may lessen activation of NADPH oxidase in various tissues by decreasing the stimulation of protein kinase C via newly synthesized diacylglycerol.[29, 53] And the lower circulating concentration of LDL particles associated with such diets could be expected to decrease NADPH oxidase-mediated endothelial oxidative stress.[54] Hence, plant-based diets, especially those that are phytochemical rich, may work in several complementary ways to minimize oxidative stress, globally or in specific tissues.

A recent clinical study shows that serum markers of oxidative stress improve in volunteers consuming a low-glycemic-index vegan diet for 21 days.[55] While this finding is consistent with the hypothesis

presented here, it is conceivable that increased phytochemical intakes are largely responsible for this finding. Hence, this hypothesis might best be tested by measuring activities of mitochondrial antioxidant enzymes in accessible tissues, such as peripheral blood leukocytes, during such a diet.

These considerations suggest that some of the beneficial health impacts of vegan diets and low insulin lifestyles may be mediated by improved control of oxidant stress. Conceivably, this phenomenon could contribute to the considerably reduced risks for “Western” cancers and cardiovascular disease typically observed in Third World cultures whose members consume quasi-vegan diets and tend to be lean. Conversely, the elevated risks for these disorders in obese individuals may be attributable in part to increased oxidative stress. Vegan diets and low-insulin lifestyles may have utility as adjuvant strategies when nutraceutical or pharmaceutical antioxidants are employed to prevent or treat disorders driven to some degree by oxidative stress.

The transcriptional activity of FOXO factors, while inhibited by Akt, is boosted by phosphorylations mediated by AMPK, and by de-acetylation via Sirt1.[56-60] Hence, measures which activate AMPK and/or Sirt1 may be useful for amplifying the impact of vegan diets or low insulin lifestyles on antioxidant defenses. Metformin presumably would be a two-edged sword in this regard, as it is thought to somehow activate AMPK by *increasing* complex I superoxide production.[61, 62] But other strategies for activating AMPK – such as vinegar, short-chain fatty acids, lipoic acid[63-70] – might be more beneficial in this regard, and effective Sirt1 activators (perhaps resveratrol or quercetin, for example[71-73]) might likewise promote FOXO-mediated antioxidant defenses.

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References

[1]. Ogg S, Paradis S, Gottlieb S, Patterson GI, Lee L, Tissenbaum HA, Ruvkun G. The Fork head transcription factor DAF-16 transduces insulin-like metabolic and

- longevity signals in *C. elegans*. *Nature* 1997 October 30;389(6654):994-9.
- [2]. Baumeister R, Schaffitzel E, Hertweck M. Endocrine signaling in *Caenorhabditis elegans* controls stress response and longevity. *J Endocrinol* 2006 August;190(2):191-202.
- [3]. Murphy CT. The search for DAF-16/FOXO transcriptional targets: approaches and discoveries. *Exp Gerontol* 2006 October;41(10):910-21.
- [4]. Willcox BJ, Donlon TA, He Q, Chen R, Grove JS, Yano K, Masaki KH, Willcox DC, Rodriguez B, Curb JD. FOXO3A genotype is strongly associated with human longevity. *Proc Natl Acad Sci U S A* 2008 September 16;105(37):13987-92.
- [5]. Flachsbarth F, Caliebe A, Kleindorp R, Blanche H, von Eller-Eberstein H, Nikolaus S, Schreiber S, Nebel A. Association of FOXO3A variation with human longevity confirmed in German centenarians. *Proc Natl Acad Sci U S A* 2009 February 24;106(8):2700-5.
- [6]. Anselmi CV, Malovini A, Roncarati R, Novelli V, Villa F, Condorelli G, Bellazzi R, Puca AA. Association of the FOXO3A locus with extreme longevity in a southern Italian centenarian study. *Rejuvenation Res* 2009 April;12(2):95-104.
- [7]. Li Y, Wang WJ, Cao H, Lu J, Wu C, Hu FY, Guo J, Zhao L, Yang F, Zhang YX, Li W, Zheng GY, Cui H, Chen X, Zhu Z, He H, Dong B, Mo X, Zeng Y, Tian XL. Genetic association of FOXO1A and FOXO3A with longevity trait in Han Chinese populations. *Hum Mol Genet* 2009 December 15;18(24):4897-904.
- [8]. Kops GJ, Dansen TB, Polderman PE, Saarloos I, Wirtz KW, Coffey PJ, Huang TT, Bos JL, Medema RH, Burgering BM. Forkhead transcription factor FOXO3a protects quiescent cells from oxidative stress. *Nature* 2002 September 19;419(6904):316-21.
- [9]. Li M, Chiu JF, Mossman BT, Fukagawa NK. Down-regulation of manganese-superoxide dismutase through phosphorylation of FOXO3a by Akt in explanted vascular smooth muscle cells from old rats. *J Biol Chem* 2006 December 29;281(52):40429-39.
- [10]. Chiribau CB, Cheng L, Cucoranu IC, Yu YS, Clempus RE, Sorescu D. FOXO3A regulates peroxiredoxin III expression in human cardiac fibroblasts. *J Biol Chem* 2008 March 28;283(13):8211-7.
- [11]. Li M, Chiu JF, Gagne J, Fukagawa NK. Age-related differences in insulin-like growth factor-1 receptor signaling regulates Akt/FOXO3a and ERK/Fos pathways in vascular smooth muscle cells. *J Cell Physiol* 2008 November;217(2):377-87.
- [12]. Tan WQ, Wang K, Lv DY, Li PF. Foxo3a inhibits cardiomyocyte hypertrophy through transactivating catalase. *J Biol Chem* 2008 October 31;283(44):29730-9.
- [13]. Olmos Y, Valle I, Borniquel S, Tierrez A, Soria E, Lamas S, Monsalve M. Mutual dependence of Foxo3a and PGC-1alpha in the induction of oxidative stress genes. *J Biol Chem* 2009 May 22;284(21):14476-84.
- [14]. Salih DA, Brunet A. FoxO transcription factors in the maintenance of cellular homeostasis during aging. *Curr Opin Cell Biol* 2008 April;20(2):126-36.
- [15]. Ouyang W, Li MO. Foxo: in command of T lymphocyte homeostasis and tolerance. *Trends Immunol* 2011 January;32(1):26-33.
- [16]. Morris BJ. A forkhead in the road to longevity: the molecular basis of lifespan becomes clearer. *J Hypertens* 2005 July;23(7):1285-309.
- [17]. Van Der Heide LP, Jacobs FM, Burbach JP, Hoekman MF, Smidt MP. FoxO6 transcriptional activity is regulated by Thr26 and Ser184, independent of nucleo-cytoplasmic shuttling. *Biochem J* 2005 November 1;391(Pt 3):623-9.
- [18]. Allen NE, Appleby PN, Davey GK, Key TJ. Hormones and diet: low insulin-like growth factor-I but normal bioavailable androgens in vegan men. *Br J Cancer* 2000 July;83(1):95-7.
- [19]. McCarty MF. Hepatic monitoring of essential amino acid availability may regulate IGF-I activity, thermogenesis, and fatty acid oxidation/synthesis. *Med Hypotheses* 2001 February;56(2):220-4.
- [20]. Allen NE, Appleby PN, Davey GK, Key TJ, Rinaldi S, Kaaks R. The effect of diet on serum insulin-like growth-factor-I and its main binding proteins. *IARC Sci Publ* 2002;156:295-6.
- [21]. Allen NE, Appleby PN, Davey GK, Kaaks R, Rinaldi S, Key TJ. The associations of diet with serum insulin-like growth factor I and its main binding proteins in 292 women meat-eaters, vegetarians, and vegans. *Cancer Epidemiol Biomarkers Prev* 2002 November;11(11):1441-8.
- [22]. Barnard RJ, Ngo TH, Leung PS, Aronson WJ, Golding LA. A low-fat diet and/or strenuous exercise alters the IGF axis in vivo and reduces prostate tumor cell growth in vitro. *Prostate* 2003 August 1;56(3):201-6.
- [23]. Ngo TH, Barnard RJ, Leung PS, Cohen P, Aronson WJ. Insulin-like growth factor I (IGF-I) and IGF binding protein-1 modulate prostate cancer cell growth and apoptosis: possible mediators for the effects of diet and exercise on cancer cell survival. *Endocrinology* 2003 June;144(6):2319-24.
- [24]. Fontana L, Klein S, Holloszy JO. Long-term low-protein, low-calorie diet and endurance exercise modulate metabolic factors associated with cancer risk. *Am J Clin Nutr* 2006 December;84(6):1456-62.
- [25]. Fontana L, Weiss EP, Villareal DT, Klein S, Holloszy JO. Long-term effects of calorie or protein restriction on serum IGF-1 and IGFBP-3 concentration in humans. *Aging Cell* 2008 October;7(5):681-7.
- [26]. Campbell TC. Dietary protein, growth factors, and cancer. *Am J Clin Nutr* 2007 June;85(6):1667.
- [27]. Lee PD, Conover CA, Powell DR. Regulation and function of insulin-like growth factor-binding protein-1. *Proc Soc Exp Biol Med* 1993 October;204(1):4-29.
- [28]. Frystyk J, Hussain M, Skjaerbaek C, Schmitz O, Christiansen JS, Froesch ER, Orskov H. Serum free IGF-I

- during a hyperinsulinemic clamp following 3 days of administration of IGF-I vs. saline. *Am J Physiol* 1997 September;273(3 Pt 1):E507-E513.
- [29]. McCarty MF. Dietary saturate/unsaturate ratio as a determinant of adiposity. *Med Hypotheses* 2010 July;75(1):14-6.
- [30]. Johnson JB, Laub DR, John S. The effect on health of alternate day calorie restriction: eating less and more than needed on alternate days prolongs life. *Med Hypotheses* 2006;67(2):209-11.
- [31]. Johnson JB, Summer W, Cutler RG, Martin B, Hyun DH, Dixit VD, Pearson M, Nassar M, Telljohann R, Maudsley S, Carlson O, John S, Laub DR, Mattson MP. Alternate day calorie restriction improves clinical findings and reduces markers of oxidative stress and inflammation in overweight adults with moderate asthma. *Free Radic Biol Med* 2007 March 1;42(5):665-74.
- [32]. Anson RM, Guo Z, de CR, Iyun T, Rios M, Hagepanos A, Ingram DK, Lane MA, Mattson MP. Intermittent fasting dissociates beneficial effects of dietary restriction on glucose metabolism and neuronal resistance to injury from calorie intake. *Proc Natl Acad Sci U S A* 2003 May 13;100(10):6216-20.
- [33]. Mattson MP. The need for controlled studies of the effects of meal frequency on health. *Lancet* 2005 June 4;365(9475):1978-80.
- [34]. Heller RF, Heller RF. Hyperinsulinemic obesity and carbohydrate addiction: the missing link is the carbohydrate frequency factor. *Med Hypotheses* 1994 May;42(5):307-12.
- [35]. Nilsson M, Holst JJ, Bjorck IM. Metabolic effects of amino acid mixtures and whey protein in healthy subjects: studies using glucose-equivalent drinks. *Am J Clin Nutr* 2007 April;85(4):996-1004.
- [36]. Melnik BC. Milk--the promoter of chronic Western diseases. *Med Hypotheses* 2009 June;72(6):631-9.
- [37]. Jenkins DJ, Kendall CW, Josse AR, Salvatore S, Brighenti F, Augustin LS, Ellis PR, Vidgen E, Rao AV. Almonds decrease postprandial glycemia, insulinemia, and oxidative damage in healthy individuals. *J Nutr* 2006 December;136(12):2987-92.
- [38]. Doi K. Effect of konjac fibre (glucomannan) on glucose and lipids. *Eur J Clin Nutr* 1995 October;49 Suppl 3:S190-S197.
- [39]. Johnston CS, Gaas CA. Vinegar: medicinal uses and antiglycemic effect. *MedGenMed* 2006;8(2):61.
- [40]. Ghiselli A, D'Amicis A, Giacosa A. The antioxidant potential of the Mediterranean diet. *Eur J Cancer Prev* 1997 March;6 Suppl 1:S15-S19.
- [41]. Kazimirova A, Barancokova M, Volkovova K, Staruchova M, Krajcovicova-Kudlackova M, Wsolova L, Collins AR, Dusinska M. Does a vegetarian diet influence genomic stability? *Eur J Nutr* 2004 February;43(1):32-8.
- [42]. Szeto YT, Kwok TC, Benzie IF. Effects of a long-term vegetarian diet on biomarkers of antioxidant status and cardiovascular disease risk. *Nutrition* 2004 October;20(10):863-6.
- [43]. Colombo C, Muti P, Pala V, Cavalleri A, Venturelli E, Locardi M, Berrino F, Secreto G. Plant-based diet, serum fatty acid profile, and free radicals in postmenopausal women: the diet and androgens (DIANA) randomized trial. *Int J Biol Markers* 2005 July;20(3):169-76.
- [44]. Krajcovicova-Kudlackova M, Valachovicova M, Paukova V, Dusinska M. Effects of diet and age on oxidative damage products in healthy subjects. *Physiol Res* 2008;57(4):647-51.
- [45]. Surh YJ, Kundu JK, Na HK. Nrf2 as a master redox switch in turning on the cellular signaling involved in the induction of cytoprotective genes by some chemopreventive phytochemicals. *Planta Med* 2008 October;74(13):1526-39.
- [46]. Richie JP, Jr., Leutzinger Y, Parthasarathy S, Malloy V, Orentreich N, Zimmerman JA. Methionine restriction increases blood glutathione and longevity in F344 rats. *FASEB J* 1994 December;8(15):1302-7.
- [47]. Orentreich N, Matias JR, DeFelice A, Zimmerman JA. Low methionine ingestion by rats extends life span. *J Nutr* 1993 February;123(2):269-74.
- [48]. Sun L, Sadighi Akha AA, Miller RA, Harper JM. Life-span extension in mice by preweaning food restriction and by methionine restriction in middle age. *J Gerontol A Biol Sci Med Sci* 2009 July;64(7):711-22.
- [49]. Lopez-Torres M, Barja G. Lowered methionine ingestion as responsible for the decrease in rodent mitochondrial oxidative stress in protein and dietary restriction possible implications for humans. *Biochim Biophys Acta* 2008 November;1780(11):1337-47.
- [50]. Caro P, Gomez J, Sanchez I, Naudi A, Ayala V, Lopez-Torres M, Pamplona R, Barja G. Forty percent methionine restriction decreases mitochondrial oxygen radical production and leak at complex I during forward electron flow and lowers oxidative damage to proteins and mitochondrial DNA in rat kidney and brain mitochondria. *Rejuvenation Res* 2009 December;12(6):421-34.
- [51]. Gomez J, Caro P, Sanchez I, Naudi A, Jove M, Portero-Otin M, Lopez-Torres M, Pamplona R, Barja G. Effect of methionine dietary supplementation on mitochondrial oxygen radical generation and oxidative DNA damage in rat liver and heart. *J Bioenerg Biomembr* 2009 June;41(3):309-21.
- [52]. McCarty MF, Barroso-Aranda J, Contreras F. The low-methionine content of vegan diets may make methionine restriction feasible as a life extension strategy. *Med Hypotheses* 2009 February;72(2):125-8.
- [53]. Inoguchi T, Li P, Umeda F, Yu HY, Kakimoto M, Imamura M, Aoki T, Etoh T, Hashimoto T, Naruse M, Sano H, Utsumi H, Nawata H. High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C--dependent activation of

- NAD(P)H oxidase in cultured vascular cells. *Diabetes* 2000 November;49(11):1939-45.
- [54]. O'Donnell RW, Johnson DK, Ziegler LM, DiMattina AJ, Stone RI, Holland JA. Endothelial NADPH oxidase: mechanism of activation by low-density lipoprotein. *Endothelium* 2003;10(6):291-7.
- [55]. Bloomer RJ, Kabir MM, Trepanowski JF, Canale RE, Farney TM. A 21 day Daniel Fast improves selected biomarkers of antioxidant status and oxidative stress in men and women. *Nutr Metab (Lond)* 2011;8:17.
- [56]. Greer EL, Oskoui PR, Banko MR, Maniar JM, Gygi MP, Gygi SP, Brunet A. The energy sensor AMP-activated protein kinase directly regulates the mammalian FOXO3 transcription factor. *J Biol Chem* 2007 October 12;282(41):30107-19.
- [57]. Greer EL, Banko MR, Brunet A. AMP-activated protein kinase and FoxO transcription factors in dietary restriction-induced longevity. *Ann N Y Acad Sci* 2009 July;1170:688-92.
- [58]. Colombo SL, Moncada S. AMPK α 1 regulates the antioxidant status of vascular endothelial cells. *Biochem J* 2009 July 15;421(2):163-9.
- [59]. Kobayashi Y, Furukawa-Hibi Y, Chen C, Horio Y, Isobe K, Ikeda K, Motoyama N. SIRT1 is critical regulator of FOXO-mediated transcription in response to oxidative stress. *Int J Mol Med* 2005 August;16(2):237-43.
- [60]. An BS, Tavera-Mendoza LE, Dimitrov V, Wang X, Calderon MR, Wang HJ, White JH. Stimulation of Sirt1-regulated FoxO protein function by the ligand-bound vitamin D receptor. *Mol Cell Biol* 2010 October;30(20):4890-900.
- [61]. Zou MH, Kirkpatrick SS, Davis BJ, Nelson JS, Wiles WG, Schlattner U, Neumann D, Brownlee M, Freeman MB, Goldman MH. Activation of the AMP-activated protein kinase by the anti-diabetic drug metformin in vivo. Role of mitochondrial reactive nitrogen species. *J Biol Chem* 2004 October 15;279(42):43940-51.
- [62]. Fujita Y, Hosokawa M, Fujimoto S, Mukai E, Abudukadier A, Obara A, Ogura M, Nakamura Y, Toyoda K, Nagashima K, Seino Y, Inagaki N. Metformin suppresses hepatic gluconeogenesis and lowers fasting blood glucose levels through reactive nitrogen species in mice. *Diabetologia* 2010 July;53(7):1472-81.
- [63]. Kondo T, Kishi M, Fushimi T, Kaga T. Acetic acid upregulates the expression of genes for fatty acid oxidation enzymes in liver to suppress body fat accumulation. *J Agric Food Chem* 2009 July 8;57(13):5982-6.
- [64]. Sakakibara S, Yamauchi T, Oshima Y, Tsukamoto Y, Kadowaki T. Acetic acid activates hepatic AMPK and reduces hyperglycemia in diabetic KK-A(y) mice. *Biochem Biophys Res Commun* 2006 June 2;344(2):597-604.
- [65]. Sakakibara S, Murakami R, Takahashi M, Fushimi T, Murohara T, Kishi M, Kajimoto Y, Kitakaze M, Kaga T. Vinegar intake enhances flow-mediated vasodilatation via upregulation of endothelial nitric oxide synthase activity. *Biosci Biotechnol Biochem* 2010;74(5):1055-61.
- [66]. Peng L, Li ZR, Green RS, Holzman IR, Lin J. Butyrate enhances the intestinal barrier by facilitating tight junction assembly via activation of AMP-activated protein kinase in Caco-2 cell monolayers. *J Nutr* 2009 September;139(9):1619-25.
- [67]. Hu GX, Chen GR, Xu H, Ge RS, Lin J. Activation of the AMP activated protein kinase by short-chain fatty acids is the main mechanism underlying the beneficial effect of a high fiber diet on the metabolic syndrome. *Med Hypotheses* 2010 January;74(1):123-6.
- [68]. Lee WJ, Song KH, Koh EH, Won JC, Kim HS, Park HS, Kim MS, Kim SW, Lee KU, Park JY. Alpha-lipoic acid increases insulin sensitivity by activating AMPK in skeletal muscle. *Biochem Biophys Res Commun* 2005 July 8;332(3):885-91.
- [69]. Lee WJ, Lee IK, Kim HS, Kim YM, Koh EH, Won JC, Han SM, Kim MS, Jo I, Oh GT, Park IS, Youn JH, Park SW, Lee KU, Park JY. Alpha-lipoic acid prevents endothelial dysfunction in obese rats via activation of AMP-activated protein kinase. *Arterioscler Thromb Vasc Biol* 2005 December;25(12):2488-94.
- [70]. Park KG, Min AK, Koh EH, Kim HS, Kim MO, Park HS, Kim YD, Yoon TS, Jang BK, Hwang JS, Kim JB, Choi HS, Park JY, Lee IK, Lee KU. Alpha-lipoic acid decreases hepatic lipogenesis through adenosine monophosphate-activated protein kinase (AMPK)-dependent and AMPK-independent pathways. *Hepatology* 2008 November;48(5):1477-86.
- [71]. Knutson MD, Leeuwenburgh C. Resveratrol and novel potent activators of SIRT1: effects on aging and age-related diseases. *Nutr Rev* 2008 October;66(10):591-6.
- [72]. Davis JM, Murphy EA, Carmichael MD, Davis B. Quercetin increases brain and muscle mitochondrial biogenesis and exercise tolerance. *Am J Physiol Regul Integr Comp Physiol* 2009 April;296(4):R1071-R1077.
- [73]. Chung S, Yao H, Caito S, Hwang JW, Arunachalam G, Rahman I. Regulation of SIRT1 in cellular functions: role of polyphenols. *Arch Biochem Biophys* 2010 September 1;501(1):79-90.